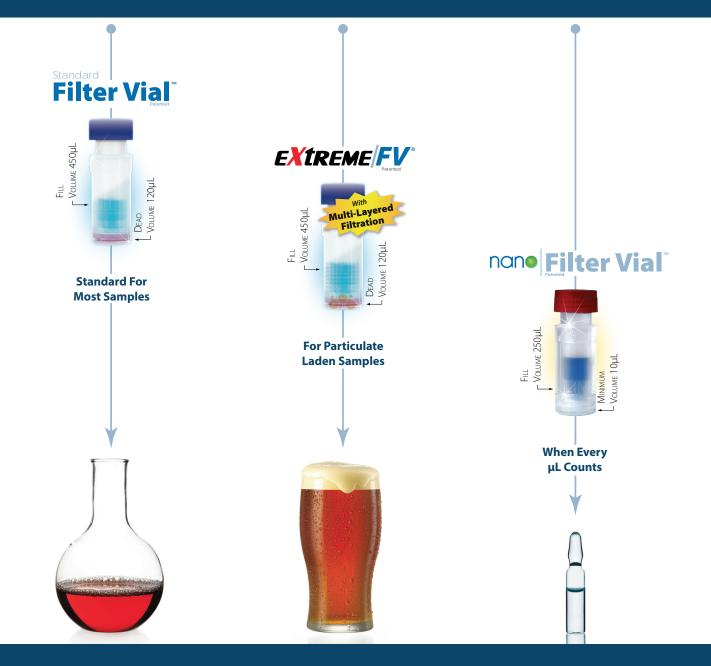


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Filter Vials have the following patents: US 7,790,117, 8,211,384, 8,383,006, 8,322,539, EU patent 2268252, EP2268252B1, Singapore Patent 164909, Worldwide Patents Pending

For up to date patent and trademark information please see htslabs.com.

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eXtreme FV® Improved Sample Preparation Methods for Athlete Doping Analysis of Common Compounds in Urine by LCMS Pesticide Applilcations - Soil & Vegitation eXtreme FV® vs SPE for the Analysis of Pesticides in Orange Juice Supplement Analysis of Huperzine A by HPLC Antibody Analysis with eXtreme FV® Analysis of Nitrosamines in Tobacco	15 16 21 23 28 29 30
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Filter Vial Overview

Thomson Filter Vials (patented) are a single system which replaces HPLC Vials, HPLC Caps, Syringes, & Syringe Filters for the filtration of samples. In 15 seconds, Filter Vials allow for Sample Preparation of unfiltered samples to filtered samples in an autosampler ready vial. The filter vial consists of two parts: a filter vial shell and a plunger which includes a filter on one end and a vial cap on the other end. Samples are filtered by pipetting the sample into the filter vial shell, inserting the plunger into the shell, and then pushing the plunger into the shell.

Thomson Filter Vials simplify general filtration by replacing syringes & syringe filters, centrifuging spin columns and/or liquid-liquid extractions.

Applications for Thomson Filter Vials include all sample types to be analyzed by HPLC, UHPLC, LC-MS and GC-MS.



 Max Fill Vol.
 450μL

 Dead Vol.
 120μL





 Max Fill Vol.
 450μL

 Dead Vol.
 120μL

Standard Filter Vials (120µL Dead Volume)

Thomson Standard Filter (*patented*) can be used for samples containing less than 10% solid particulates. The filter vial consists of two parts: a filter vial shell and a plunger which includes a single layer filter on one end and a vial cap on the other end.

Applications for Thomson Standard Filter Vials include filtration of catalysts from organic and medicinal chemistry synthesis reactions, saccharide analysis in corn syrup, and in-vial protein precipitation.

eXtreme[**FV**[®] (Multi-Layered Filtration)

Thomson eXtreme|FV[®] (*patented*) offer multi-layer filtration for viscous samples and samples containing up to 30% solid particulates. The filter vial consists of two parts: a filter vial shell and a plunger which includes a multi-layer filter on one end and a vial cap on the other end.

eXtreme|FV[®] allow for compounds to be separated from the matrix which, results in both a higher signal to noise ratio and peaks that are more differentiated.

Prior to the introduction of the eXtreme|FV[®], many samples containing high levels of particulates were "filtered" by using an SPE step in the method. These methods are readily amendable to the replacement of the SPE step using a rapid and lower cost eXtreme|FV[®] step.

Applications for Thomson eXtreme|FV[®] include filtration of cell and cell debris from cell culture; pesticide analysis in food, tissue, soil, and water; and toxicology analysis in blood and urine.



Max Fill Vol.250μLMin Fill Vol.10μL (for 2μL injection)

nano|**Filter Vials**[™] (10µL Minimum Volume)

Thomson nano|Filter Vials^m offer a very low dead volume allowing one to filter as little as 10µL of sample with enough remaining filtrate to make a 2µL injection. The filter vial consists of two parts: a filter vial shell with mating bottom surface and a plunger which includes a filter on one end and a screw cap vial on the other end.

Applications include the analysis of enzymes, peptides, DNA, RNA, synthesis reaction intermediates, finished products, saliva, samples available in low volumes, in-vial evaporation and re-suspension for sample concentration and buffer/solvent change.

Application Selection



____ ____

See our Technical Lik for more Application	orary Is	^{nah}	Stan Persit	Low Startd Filter L	ets. etsologion	eme/FV.
htslabs.com	1ªr	1ªL	es s	107		
10µL-100µL						
120µL-450µL						
UPLC Compatible						
GCMS Compatible						
≤ 30% Solids						
Viscous						
Replacement for SPE						
General Liquids < 10% solids						
Cell Fermentation						
Particulate Removal						
Automation Compatible						
Small Molecules						
Food & Supplements						
Toxicology						
Pesticides						
Environmental						
Sterile Testing						

Filter Vial Membrane Material

The recommended membrane for sample filtration is based on the percentage of organic solvent in the sample and the amount of protein binding.

	AQUEOUS	ORGANIC	LOW PROTEIN BINDING
PTFE			
PVDF	\diamond		<pre> </pre>
Nylon	6		
PES	Ô		8

Filter Vial Membrane Pore Size

The recommended membrane pore size for sample filtration is based on the cell or cell debris content of the sample and the particle size of the packing material in the chromatography column used to analyze the sample. If the sample contains cells or cellular debris, then a $0.2\mu m$ pore size membrane is recommended to maintain system sterility.

Vial Mambrana Para Siza
Add membrane pore size for sample filtration is ell or cell debris content of the sample and the the packing material in the chromatography column e the sample. If the sample contains cells or cellular 0.2µm pore size membrane is recommended to m sterility.
0.2µm Pore Size
0.45µm Pore Size

Filter Vial Leachables

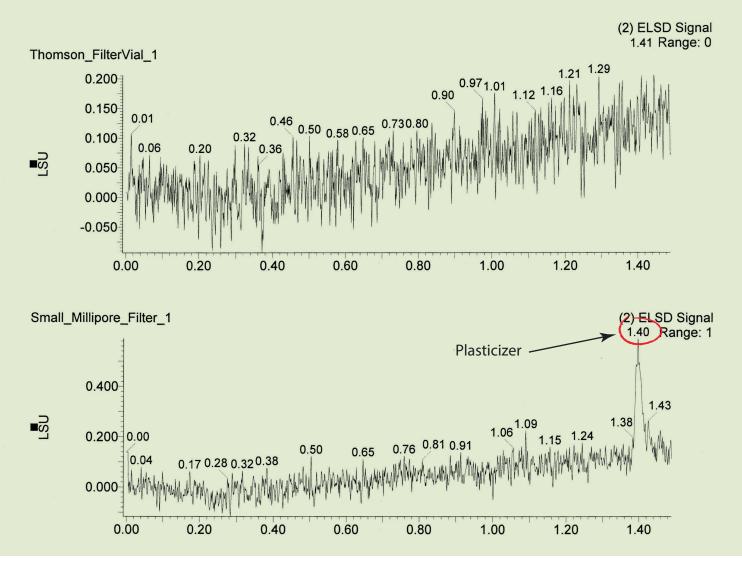
Thomson Filter Vials are manufactured without the use of plastizers or mold release agents making them LC/MS clean.

Testing with ELSD, PDA, and MS detection by Takeda Pharmaceutical showed no leaching from Thomson Standard Filter Vial with a 0.45um, PTFE membrane compared to significant leaching from Millipore Millex-FH[®] Filter, 0.45µM, hydrophobic PTFE, 4mm.

Method: A. Water B. ACN 45-90% with .05% TFA Ballistic Gradient over 1.4 minutes using Waters® Acquity® UPLC Thomson Filter Vial (patented) Part # 35540-500 Filter Vial 0.45µM hydrophobic PTFE, w/ Pre-Slit Cap Millipore Syringe Filter Part #:SLFHR04NL Millex-FH® Filter, 0.45µM, hydrophobic PTFE, 4mm, non-sterile

Plasticizer Leachable

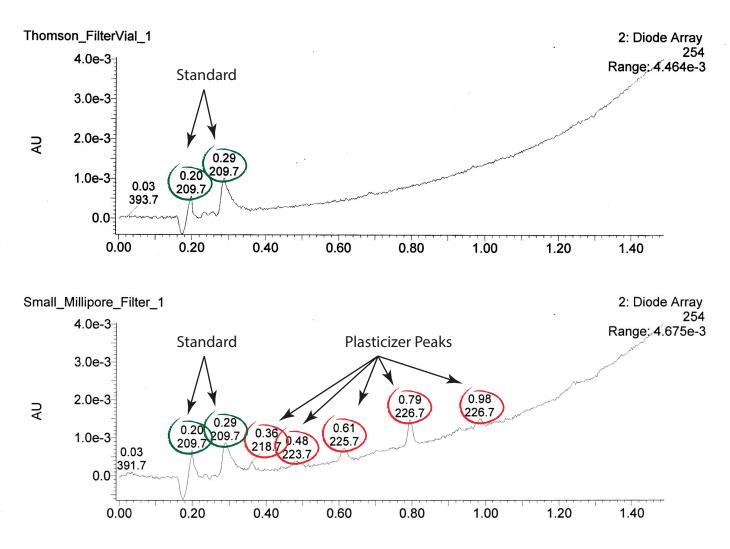
Testing by Takeda® Mass Spec. ES+ Data



Method: A. Water B. ACN 45-90% with .05% TFA Ballistic Gradient over 1.4 minutes using Waters® Acquity® UPLC Thomson Filter Vial (patented) Part # 35540-500 Filter Vial 0.45uM hydrophobic PTFE, w/ Pre-Slit Cap Millipore Syringe Filter Part #:SLFHR04NL Millex-FH® Filter, 0.45 µm, hydrophobic PTFE, 4 mm, non-sterile.

Plasticizer Leachable

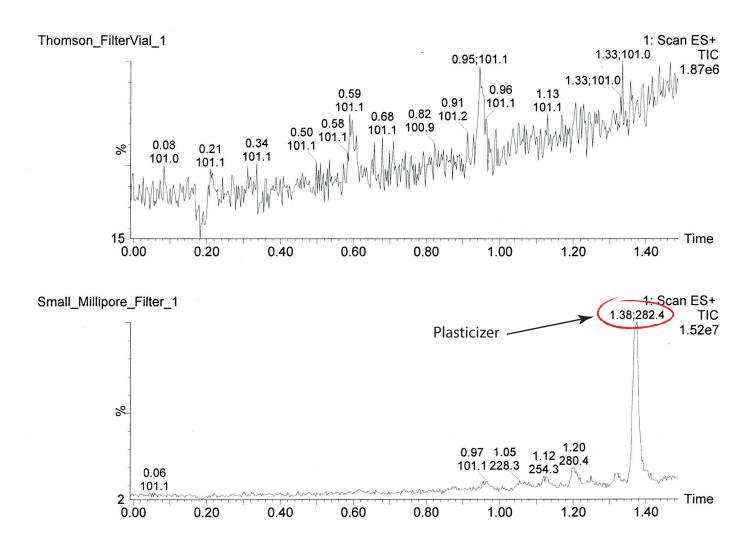
Testing by Takeda® UV Data



Method: A. Water B. ACN 45-90% with .05% TFA Ballistic Gradient over 1.4 minutes using Waters® Acquity® UPLC Thomson Filter Vial (patented) Part # 35540-500 Filter Vial 0.45uM hydrophobic PTFE, w/ Pre-Slit Cap Millipore Syringe Filter Part #:SLFHR04NL Millex-FH® Filter, 0.45 µm, hydrophobic PTFE, 4 mm, non-sterile.

Plasticizer Leachable

Testing by Takeda[®] Mass Spec ES+ Data Part 2



Method: A. Water B. ACN 45-90% with .05% TFA Ballistic Gradient over 1.4 minutes using Waters® Acquity® UPLC Thomson Filter Vial (patented) Part # 35540-500 Filter Vial 0.45uM hydrophobic PTFE, w/ Pre-Slit Cap Millipore Syringe Filter Part #:SLFHR04NL Millex-FH® Filter, 0.45 µm, hydrophobic PTFE, 4 mm, non-sterile.

Standard Filter Via

Standard For Most Samples

DEAD - VOLUME 120µL

VOLUME 450µL

FIL

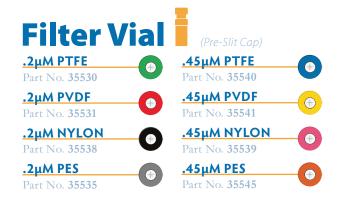
Standard Filter Vials (120µL Dead Volume)

Thomson Standard Filter (*patented*) can be used for samples containing less than 10% solid particulates. The filter vial consists of two parts: a filter vial shell and a plunger which includes a single layer filter on one end and a vial cap on the other end.

ТМ

Applications for Thomson Standard Filter Vials include filtration of catalysts from organic and medicinal chemistry synthesis reactions, saccharide analysis in corn syrup, and in-vial protein precipitation.







SINGLE SIEP[®] Filter Vials Patented Open Access LCMS

















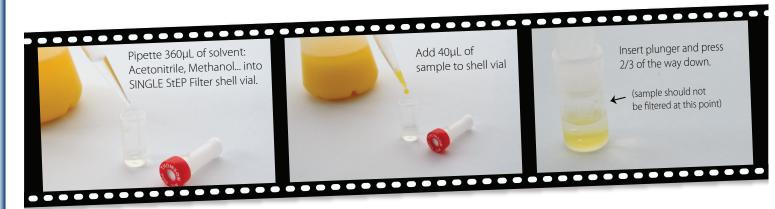
"We've been pounding our walk up systems for **over a year** without a single clog."

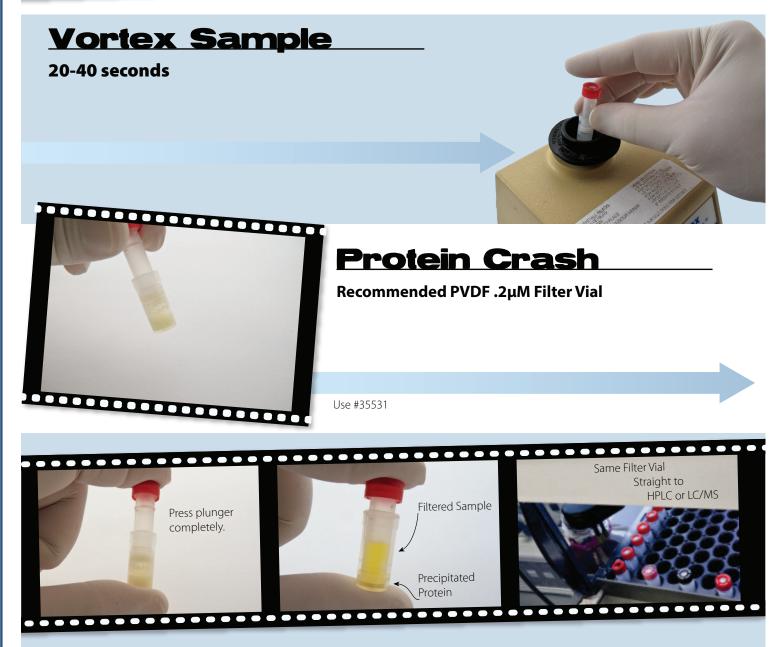
> -Justin University of Arizona

Thomson Instrument Company is not affiliated with Waters® its product the UPLC® Aquity® or The University of Arizona.

Filter Vial

Protein Precipitation with SINGLE StEP Filter Vials[™] Patented







Filter Vial

How to Mass Spec Your TLC Spots Using Thomson Filter Vials

 After eluting analytical TLC plates, scrape-off desired spots into Thomson Shell Vial





2. Add 0.4 mL EtOAc to Shell Vial

3. Next, insert Plunger, press half way down and shake or swirl to extract compound from silica gel.





4. Press down plunger completely to filter sample

5. Same Filter Vial

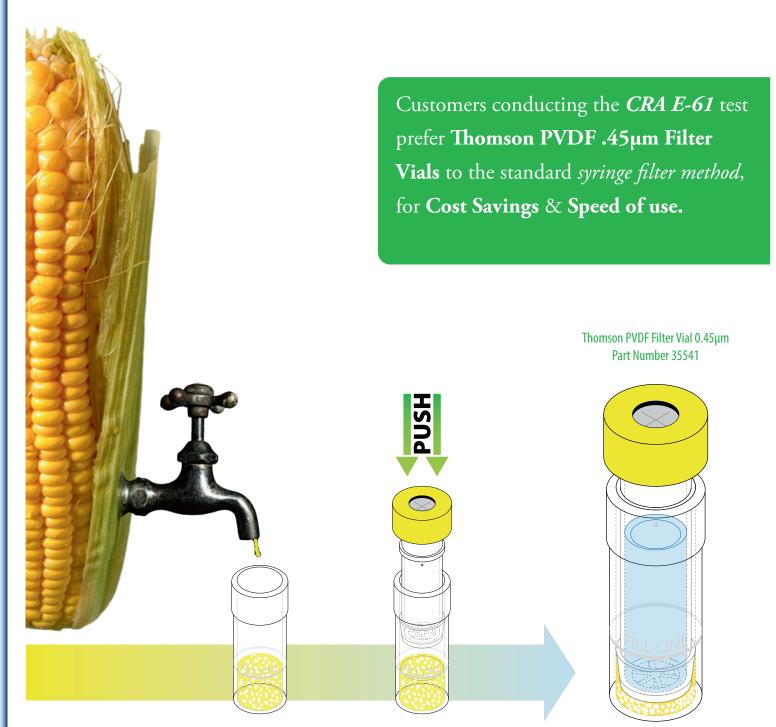
straight to Mass Spec

Filter Vial

CORN SYRUP ANALYSIS SACCHARIDES (LIQUID CHROMATOGRAPHY) with Filter Vials



Patented



*Thomson Instrument Company is not affiliated with Corn Refiners Association, Inc. and this system is not sanctioned by Corn Refiners Association, Inc., although some analysts are using Filter Vials in the Field for this purpose.





For Particulate Laden Samples

With Multi-Layered

Filtration

VOLUME 120µ

DEAD

Volume 450µl

FILL

eXtreme FV (Multi-Layered Filtration)

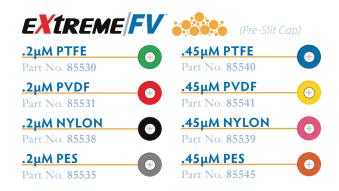
Thomson eXtreme|FV[®] (*patented*) offer multi-layer filtration for viscous samples and samples containing up to 30% solid particulates. The filter vial consists of two parts: a filter vial shell and a plunger which includes a multi-layer filter on one end and a vial cap on the other end.

eXtreme|FV[®] allow for compounds to be separated from the matrix which, results in both a higher signal to noise ratio and peaks that are more differentiated.

Prior to the introduction of the eXtreme |FV[®], many samples containing high levels of particulates were "filtered" by using an SPE step in the method. These methods are readily amendable to the replacement of the SPE step using a rapid and lower cost eXtreme |FV[®] step.

Applications for Thomson eXtreme|FV[®] include filtration of cell and cell debris from cell culture; pesticide analysis in food, tissue, soil, and water; and toxicology analysis in blood and urine.







Improved Sample Preparation Methods for Athlete Doping Analysis of Common Compounds in Urine by LCMS



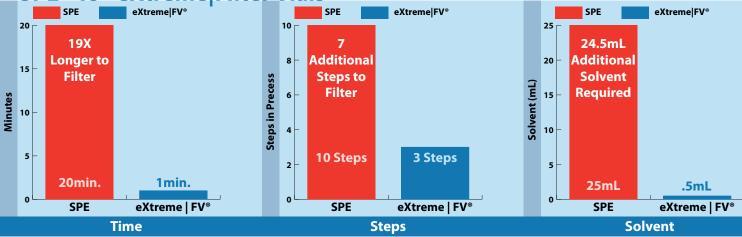
Australian Government Department of Industry



Thomson Instrument Company is not affiliated with Australian Sports Drug Testing Laboratory or World Anti-Doping Agency. World Anti-Doping Agency and Australian Sports Drug Testing Laboratory is not affiliated, nor endorses Thomson's products.

Authors : Dr. Catrin Goebel², Lisa Wanders¹, Sam Ellis¹ Thomson Instrument Company¹ Australian Sports Drug Testing Laboratory in the National Measurement Institute Department of Industry² For reprints contact folks@htslabs.com

SPE -vs- eXtreme|Filter Vials®



Abstract

Anti-doping testing by urine analysis requires fast and robust screening methods with repeatable sample preparation. Since, every sample has to be screened, methods are designed to be sufficiently sensitive and specific to identify all suspect samples. One must be careful to minimize false suspects. Ensuring samples are spiked with internal standards accordingly will help verify that samples are being extracted and tested correctly and with accurate uniformity.

The Australian Sports Drug Testing Laboratory, our collaborators, have invested time in determining a limited number of comprehensive screening methods. These methods, using Thomson's eXtreme Filter Vials (patented), comply with the World Anti-Doping Agency's (WADA) Prohibited List.

In exploring new methods labs have looked at both detection and sample prep as routes to quicker and more accurate analysis. Liquid chromatography coupled with mass spectrometry detection is prevalent, superseding many of the gas chromatographic coupled with mass spectrometry methods because of the simpler sample preparation. Specifically, the anti-doping testing



17

shown below consisted of sample preparation without the initial use of cumbersome traditional SPE methods, and instead consisted of the comparison of filtration techniques. Filter plates versus Thomson eXtreme Filter Vials (patented) were tested to determine which product allowed for a method of simple and quick urine analysis while complying with the WADA's guidelines.

Experiment

The experiments were performed at the National Measurement Institute (Australia) in the Sports Drug Testing Laboratory.

The 11.8 minute run time for the instrumental analysis meets the requirements of the WADA Technical Document- Minimum Required Performance Level (TD2013MRPL). This document details the analysis of a large number of analytes from the classes on the WADA Prohibited List, while meeting sensitivity requirements. The analytes included compounds in the following classes anabolic agents, B2-agonists, hormone antagonists and modulators, diuretics, stimulants, narcotics, glucocorticoids, B-blockers, etc.

Full Method:

A comparison between sample preparation using filter plates sourced from several different manufactures, and Thomson eXtreme Filter Vials (patented) PVDF 0.2µm (85531-500) was conducted. The preparation with the Thomson eXtreme Filter Vials were automated using a Tecan robotics platform for liquid dispensing in the Thomson 48 position rack (#35010-RACK), and 48 position press (#35010).

Direct Urine Preparation:

- 1. Label each eXtreme Filter Vial with sample/quality control sample information.
- 2. Pipette 200 μL of each sample into labeled eXtreme Filter Vial.
- 3. Add 200 µL of the Mefruside Internal Standard (300 ng/mL in 0.5% formic acid) to each filter vial cup.
- 4. Place the eXtreme Filter Vial tops onto each vial and press shut.

LCHRMS System:

UPLC coupled to High Resolution Mass Spectrometry with an electrospray source in full scan mode. Data acquisition in both positive and negative polarity modes within a single 11.8 min chromatographic run.

www.bgb-info.com

Column: C18, 2.1mm × 50mm, 1.7μm Column Temperature: 30 °C Flow rate: 300μL/min

Mobile Phase:

A: 0.3% aqueous Formic Acid in Water B: 0.3% Formic Acid in Acetonitrile

Gradient:

Time	A%	B%
0.00	95	5
0.50	95	5
3.50	80	20
5.50	75	25
7.00	43	57
8.00	10	90
8.60	10	90
8.80	95	5

Injection volume: 10μL Sample tray temperature: 18°C Column Temperature: 30°C Method run time: 11.8 minutes Gas: UHP Nitrogen



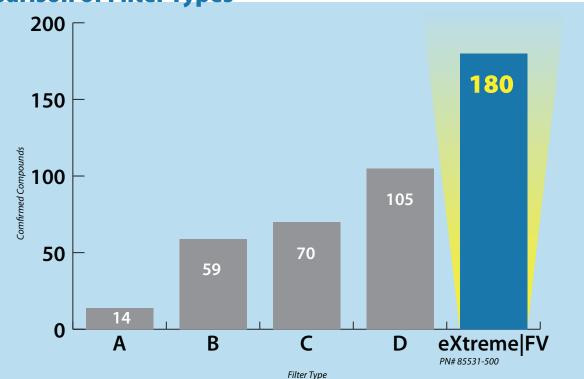
Conclusions

The Thomson eXtreme Filter Vials (patented) PVDF 0.2µm (85531-500) performed the best in compound extraction and identification while allowing the end user to follow the WADA validated method. The elimination of SPE steps from laboratory methods is a large time saver, and enables urine-direct-injection solely using Thomson eXtreme Filter Vials for filtration. Together the Thomson 48 position Filter Vial Press and automation enabled 48 position rack equaled timing of filter plate methodology but provided the best extraction and identification of all filter types. A total of 180 compounds can be identified through the screening analysis with the Thomson eXtreme Filter Vials (patented) PVDF 0.2µm (85531-500).

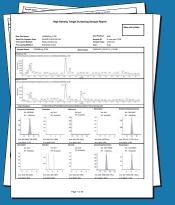
The method presented is being used for the analysis of athlete's urine samples for banned substances at the Australian Sports Drug Testing Laboratory.

Acknowledgments

We would like to thank Dr. Catrin Goebel, Director, of Australian Sports Drug Testing Laboratory in the National Measurement Institute, Department of Industry (a WADA accredited laboratory in Australia) for her extensive testing. Dr. Goebel is also an Executive member of World Association of Anti-Doping Scientist.



Comparison of Filter Types

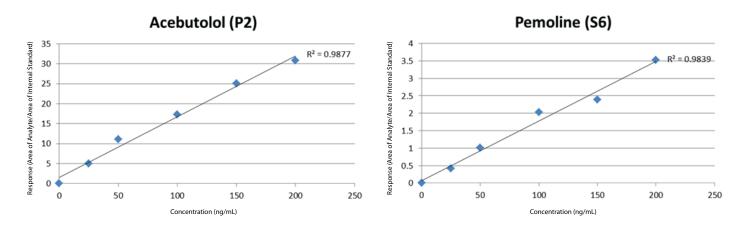


To View All Chromatograms Visit http://bit.ly/wada-data

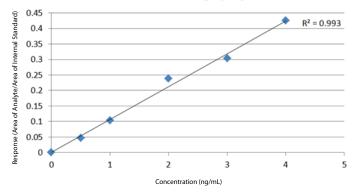




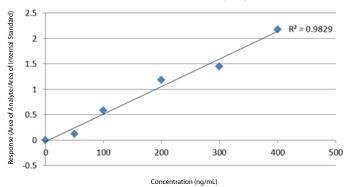
Linearity of The Analysis Method Was Assessed Over a Range From 25% To 200% Of MRPL With R2 Generally Being Greater Than 0.98



Norfentanyl (S7)



Quinethazone (S5)



Time is Equal



With automation our customers are utilizing Filter Vials at the same speed filter plates were used in the past.



Confirmed Compounds (180)

Sample Name 130524Exp_0136

Raw File 130524Exp_0136

Compound Name

State

5-Hydroxyindapamide Bisdesmethylsibutramine Desmethylsibutramine Exemestane ISD.01 Mefruside (+) ISD.02 Mefruside (-) ISD.03 D3-epitestosterone glucuronide ISD.04 D3-epitestosteronea M1.03 AICAR M1.04 GW1516 P2.03 Atenolol P2.05 Bisoprolol P2.12 Esmolol P2.14 Metipranolol P2.16 Nadolol P2.17 Nadoxolol P2.18 Oxprenolol S1.00 Clenbuterol S1.00 Gestrinone S1.00 Methyldienolone S1.00 Methyltrienolone S1.00 Metribolone S1.00 Tetrahydrogestrinone S1.00 Tibolone S1.00 Zilpaterol S1.01 3'-Hvdroxystanozolol S1.02 4'-Hydroxystanozolol S3.01 Bambuterol S3.03 Formoterol S3.04 Salbutamol S3.05 Salmeterol S3.06 Terbutaline S4.00 Andarine S4.1.00 Exemestane metabolite S4.1.01 Aminoglutethimide S4.2.00 Raloxifene S4.3.00 Fulvestrant \$4.5.00 GW1516 (501516) S5.00 Methazolamide S5.00 Piretanide S5.00 Quinethazone S5.00 Spironolactone s5.00 Trichlormethiazide S5.01 Acetazolamide S5.02 Althiazide S5.02 Amiloride S5.03 Bendroflumethiazide S5.03 Benzthiazide S5.04 Bumetanide S5.05 Canrenone S5.06 Chlorexolone S5.07 Chlorothiazide S5.08 Chlorthalidone S5.09 Clopamide S5.1.01 Probenecid S5.10 Cyclopenthiazide S5.11 Cvclothiazide S5.12 Dichlorphenamide S5.13 Epitizide S5.14 Eplenerone S5.15 Etacrynic acid (frag?)

Found Confirmed Confirmed

Batch Name 130524CG 26-05-13 141348

S5.16 Furosemide S5.17 Hydrochlorothiazide S5.20 Mefruside metabolite 2 S5.21 Indapamide S5.22 Metolazone S5.23 Polythiazide S5.24 Torasemide S5.25 Triamterene S5.26 Xipamide S6.00 Caffeine S6.00 Cis-4-Methylaminorex S6.00 Cotinine (Nicotine metab) S6.00 MBDB S6.00 Methoxyamphetamine S6.00 Methylenedioxyethylamphetamine S6.01 Adrafinil S6.03 Amiphenazole S6.04 Amphetamine S6.07 Benzoylecgonine S6.09 Benzylpiperazine S6.10 Carphedon S6.11 Cathine S6.14 Crotethamide S6.15 Cyclazodone S6.17 Ephedrine S6.17 Phenylpropanolamine S6.17 Pseudoepherine S6.18 Etamivan S6.20 Etilefrine S6.25Fenetylline S6.30 Hydroxy mesocarb S6.32 Isometheptene S6.33 Methylenedioxyamphetamine (MDA) S6.34 Methylenedioxymethylamphetamine(MDMA) S6.43 Methylphenidate S6.44 Modafinil S6.45 Modafinil Acid (metabolite) S6.46 Nikethamide S6.49 Oxilofrine S6.50 Pemoline S6.51 Pentetrazol S6.53 Phenmetrazine S6.56 Pholedrine S6.57 p-Hydroxy amphetamine S6.62 Ritalinic Acid S6.64 nor-Selegiline S7.00 Methylecoonine S7.03 Codeine S7.06 Hydromorphone S7.08 Morphine S8.04 JWH018 N-(5-hydroxypentyl) metabolite S8.05 JWH073 N-butanoic acid metabolite S9.03 Budesonide S9.05 Cortisol S9.06 Cortisone S9.12 Flumethasone S9.16 Fluticasone propionate metabolite S9.17 Methylprednisolone S9.18 16a-OH-Prednisolone S9.18 Prednisolone Sildenafil Tadalafil Vardenafil

Confirmed Confirmed







Vegetation & Soil Application

1. Samples are extracted using 20g of homogeneous, ground sample

2. Sample clean-up was achieved using Thomson eXtreme Filter Vials (PTFE .2µm & PVDF .2µm)

The following compounds were seen in both soil and vegetation:

MCPP Clopyralid Aminopyralid Picloram Dicamba

.1.

Quinclorac Fluroxypyr MCPA Diflufenzopyr

System:	UPLC [®] /MS/MS [®]
HPLC Column:	Zorbax Rx C8, 150 x 2.1 mm id
HPLC Guard Column:	Agilent Eclipse XDB-C8, 2.1 x 12.5mm, 5 micron
Column Temperature:	35°C
Autosampler Temperature:	15°C
Injection Volume:	10μl
Run Time:	8 min
Solvent A :	0.15% Glacial Acid in Water
Solvent B:	0.15% Glacial Acid in ACN

	o(B	0/ D	
Flow Rate (ml/min)	%A	%B	
0.8	95	5	
0.8	95	5	
0.8	80	20	
0.8	70	30	
0.8	60	40	
0.8	50	50	
0.8	5	95	
0.8	5	95	
0.8	95	5	
	0.8 0.8 0.8 0.8 0.8 0.8 0.8 0.8	0.8 95 0.8 95 0.8 80 0.8 70 0.8 60 0.8 50 0.8 5 0.8 5	0.89550.89550.880200.870300.860400.850500.85950.8595



Tar Veg Sj 070313_39	ok		V	eç	ge	eta	ati	io	n	TQD: G	BB814									ul-2013 23:29: of 2 Channels E 4.18 TIC (MCF A 2.32
070313_39	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80		4.20 of 2 Channels E 12 TIC (2,4 D 1.35
070313_39	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80 3.		4.20 of 2 Channels E TIC (Triclop 6.89
070313_39	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60 3.	3.80	4.00 9: MRM	4.20 of 2 Channels E TIC (MCF 2.18
0 ³	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60 3.60	3.80	4.00 8: MRM	4.20 of 2 Channels E TIC (2,4 1.38
0 ³	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20 3.13 1 3.14	3.40	3.60	3.80		4.20 of 2 Channels E TIC (Diflufenzop 9
0 070313_39 100	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80 2.85	3.00	3.20	3.40	3.60	3.80	4.00 5: MRM	4.20 of 2 Channels E TIC (Fluroxyp 4.68
0 ⁻³ 070313_39 100	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40 2.46	2.60 2.49	2.80	3.00	3.20	3.40	3.60	3.80	4.00	4.20 I of 1 Channel E TIC (Quinclora 1.70
0 ³	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20 2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80	4.00 3: MRM	4.20 of 2 Channels E TIC (Dicamb 6.20
0 ⁴	0.40	0.60 0.58 0.61	0.80	1.00	1.20	1.40	1.60	1.80		2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80	4.00	4.20 I of 1 Channel E TIC (Clopyral 5.82
0 ⁴	0.40	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80	4.00	4.20 Tim

TR Soil Spk 070313_50			S	oi					TQD: 0	QBB814	ŀ								ul-2013 01:07:48 of 2 Channels ES- 4.18 TIC (MCPP) 0.11e ²
0 ⁻³	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80		4.20 of 2 Channels ES- 12 TIC (2,4 DP) 5.02e4
0 ⁴	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80	4.00 10: MRM 88	4.20 of 2 Channels ES- TIC (Triclopyr) 1.75e4
0 ⁴	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60 3.	3.80		4.20 of 2 Channels ES- TIC (MCPA) 8.35e4
0.40 070313_50	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60 3.60	3.80	4.00	4.20 of 2 Channels ES- TIC (2,4 D) 4.65e4
0 ⁴ 0.40 070313_50 100	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20 3.13	3.40	3.60	3.80		4.20 of 2 Channels ES- TIC (Diflufenzopyr) 3.64e3
0 ³	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80 2.85	3.00	3.20	3.40	3.60	3.80	4.00	4.20 of 2 Channels ES- TIC (Fluroxypyr) 9.2563
0 ⁴		0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60 2.48	2.80	3.00	3.20	3.40	3.60	3.80		4.20 I of 1 Channel ES- TIC (Quinclorac) 6.67e3
0 ⁴		0.80	1.00		1.40	1.60	1.80	2.00	2.20 2.2	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80	4.00	4.20 of 2 Channels ES- TIC (Dicamba) 2.30e4
0 ⁻³ 0.40 070313_50	0.60 0.58	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80	4.00 1: MRN	4.20 I of 1 Channel ES- TIC (Clopyralid 2.74e4
0 ^{.4}	0.60	0.80	1.00	1.20	1.40	1.60	1.80	2.00	2.20	2.40	2.60	2.80	3.00	3.20	3.40	3.60	3.80	4.00	4.20 Time





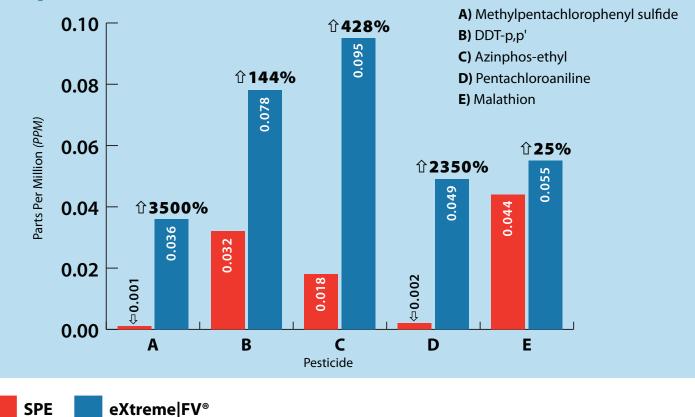
eXtreme Filter Vials® vs SPE for the analysis of Pesticides in Orange Juice



Thomson Instrument Company is not affiliated with Micro Quality Labs Inc.. Micro Quality Labs Inc. is not affiliated with Thomson Instrument Company or endorse Thomson's products.

Authors: Uday Sathe¹, Karine Aylozyan¹, Lisa Wanders², Joe Machamer², and Sam Ellis² Micro Quality Labs¹ Thomson Instrument Company² *For reprints contact folks@htslabs.com*

Comparison of Pesticide Recoveries





Abstract

Pesticides act as toxins when found in sufficient quantities as residues in food. This is of particular importance for orange juice because it is consumed in high quantities by children. Sensitive, rapid, and cost effective analytical methods are required in order to reduce the risk to consumers.

Solid Phase Extraction (*SPE*) is a common sample preparation technique used prior to GC or LC analysis of pesticides in food. Typically, SPE is used to concentrate analytes, reduce interference from co-eluting molecules or to clean up/"filter" sample particulates. Drawbacks to the use of SPE include cost, sample preparation time, large sample volumes, use and disposal of organic solvents, and potentially poor recoveries. The continuing development of higher sensitivity instrumentation and improved filtration devices has led many labs to investigate whether methods can be adapted to eliminate the SPE step.

Thomson eXtreme[®] Filter Vials offer multi-layer filtration for viscous samples and samples containing up to 30% solid particulates. Filtration time from unfiltered sample transfer to filtered sample in an autosampler ready vial is only 15 seconds. The filter vial consists of two parts: a filter vial shell and a plunger which includes the multi-layer filter on one end and a vial cap on the other end. Samples are filtered by pipetting the sample into the filter vial shell, inserting the plunger into the shell, and then pushing the plunger into the shell.

Prior to the introduction of the eXtreme Filter Vials, many samples containing high levels of particulates were only "filtered" by using an SPE step in the method. These methods are readily amendable to the replacement of the SPE step with a much faster and lower cost eXtreme Filter Vial step.

Experiment

Samples were prepared and analyzed at Micro Quality Labs, Burbank, CA.

Sample Preparation:

- 1.) Spike 10mL of commercially available High Pulp Orange Juice with 1mL of 1 ppm pesticide standard mix in a 40mL vial.
- 2.) Add one pack (approximately 6g) of Restek Extraction Salts (Restek catalog #26236) to the spiked orange juice.
- 3.) Extract the spiked orange juice with 4 x 25mL portions of methylene chloride.
- 4.) Concentrate to dryness using a Turbovap II concentrator.
- 5.) Dissolve the residue in approximately 10mL of acetonitrile.
- 6.) Vortex and sonicate the re-suspended residue with frequent swirling.
- 7.) Split the re-suspended residue into two 5mL portions.
- 8.) Dilute each 5mL portion with acetonitrile to 10mL using a volumetric flask.
- 9.) Label one flask "for SPE" and the other "for Thomson eXtreme Filter Vial".

SPE Cleanup Prior to Analysis - Restek 6mL Combo SPE Cartridge

1.) Wash one Restek 6mL Combo SPE Cartridge (*packed with 200mg CarboPrep 200 and 400mg PSA Restek catalog #26127*) with acetonitrile.

2.) Add the 10mL portion of the re-suspended residue from the flask labeled "for SPE" to the SPE cartridge.

- 3.) Elute the sample from the cartridge with 50mL of acetonitrile.
- 4.) Concentrate the eluted sample to 10mL using a Turbovap II concentrator.

Thomson eXtreme Filter Vial Cleanup Prior to Analysis

 Add 400μL of the re-suspended residue from the flask labeled "for Thomson eXtreme Filter Vial" to the shell of one Thomson eXtreme Filter Vial 0.45μm, PTFE (*Thomson Part Number 85540-500*).
 Insert plunger completely.

Analysis

Samples were analyzed utilizing an Agilent Technologies® GC/MS, 7000 Triple Quad system equipped with a 7890A GC system and 7693 auto sampler.





Compound/SAMPLE NAME	SPE+ ROUTINE Syringe FILTER	ONLY EXTREME FV W/O SPE
Alachlor	0.043	0.053
Aldrin	0.025	0.032
Azinphos-ethyl	0.018	0.095
Azinphos-methyl	0.023	0.115
BHC-alpha (benzene hexachloride)	0.026	0.033
BHC-beta	0.054	0.073
BHC-delta	0.062	0.081
BHC-gamma (Lindane, gamma HCH)	0.032	0.043
Bromophos-ethyl	0.025	0.057
Bromopropylate	0.063	0.076
Carbophenothion	0.051	0.071
Chlordane-cis (alpha)	0.04	0.052
Chlordane-oxy	0.034	0.042
Chlordane-trans (gamma)	0.039	0.049
Chlorfenvinphos	0.061	0.071
Chlorpyrifos	0.035	0.047
Chlorpyrifos-methyl	0.035	0.046
Cyfluthrin I	0.082	0.113
Cyhalothrin (lambda)	0.076	0.091
Cypermethrin I (Zeta)	0.082	0.117
Cypermethrin II {CAS # 52315-07-8}	0.08	0.113
Cypermethrin III (Beta)	0.058	0.104
Cypermethrin IV {CAS # 52315-07-8}	0.07	0.097
DCPA (Dacthal, Chlorthal-dimethyl)	0.04	0.048
DDD-o,p'	0.052	0.06
DDD-p,p'	0.056	0.066
DDE-o,p'	0.043	0.039
DDE-p,p'	0.045	0.057
DDL-p,p	0.035	0.065
DDT-p,p'	0.032	0.078
Deltamethrin	0.053	0.102
Diazinon	0.033	0.035
Dicofol		0.028
Dieldrin	0.033	
	0.041	0.052
Dimethoate	0.061	0.077
Endosulfan I (alpha isomer)	0.041	0.076
Endosulfan II (beta isomer)	0.053	0.065
Endosulfan sulfate	0.061	0.074
Endrin	0.045	0.058
Ethion	0.057	0.069
Etrimfos	0.03	0.038
Fenchlorphos oxon	0.047	0.061
Fenitrothion	0.041	0.053



	0.000	0.070
Fenpropathrin	0.068	0.078
Fensulfothion	0.1	0.117
Fenthion	0.041	0.05
Fenthion sulfone	0.081	0.107
Fenthion sulfoxide	0.106	0.134
Fenvalerate I	0.076	0.106
Fenvalerate II {CAS # 51630-58-1}	0.055	0.073
Fluvalinate-tau l	0.078	0.082
Fluvalinate-tau II {CAS # 102851-06-9}	0.058	0.084
Fonofos	0.023	0.028
Heptachlor	0.022	0.029
Heptachlor endo-epoxide (isomer A)	0.039	0.048
Heptachlor exo-epoxide (isomer B)	0.037	0.045
Hexachlorobenzene	0	0.019
Malaoxon (metabolite of Malathion)	0.07	0.086
Malathion	0.044	0.055
Mecarbam	0.052	0.062
Methidathion	0.063	0.08
Methylpentachlorophenyl sulfide	0.001	0.036
Mirex	0.042	0.056
Octachlorodipropyl ether (S421)	0.021	0.047
Omethoate	0.052	0.061
Paraoxon	0.071	0.08
Parathion	0.039	0.049
Parathion-methyl	0.035	0.045
Pendimethalin	0.038	0.048
Pentachloroaniline	0.002	0.049
Pentachloroanisole	0.017	0.021
Permethrin I	0.068	0.097
Permethrin II (trans)	0.071	0.115
Phosalone	0.005	0.089
Phosmet	0.031	0.104
Piperonyl butoxide	0.117	0.105
Pirimiphos-ethyl	0.044	0.053
Pirimiphos-methyl	0.04	0.05
Procymidone	0.064	0.082
Profenofos	0.058	0.071
Prothiofos	0.033	0.06
Quinalphos	0.042	0.061
Quintozene	0.02	0.028
Ronnel (Fenchlorphos)	0.031	0.020
Tecnazene (TCNB)	0.011	0.014
Tetradifon	0.062	0.077
Vinclozolin	0.043	0.052
VIIICIOZOIIN	0.045	0.032





GCMS Data (links to PDF)

With	Out	Internal	Snike
VVICII	out	interna	JUIKE

SPE w/ Filtration || eXtreme|FV® 85540 ||

| http://bit.ly/spe-spike | http://bit.ly/extreme-no-spike

USP 36 <561> with 0.1 PPM	http://bit.ly/usp-spike
eXtreme FV [®] with 0.1 PPM	http://bit.ly/extreme-with-spike

Conclusions

With Internal Spike

The Thomson eXtreme 0.45µm, PTFE Filter Vials patented (*Thomson #85540-500*) yielded 26% higher recoveries on average when tested with 87 common pesticides. In the cases highlighted in the results table, greater than 428% recovery increases were seen. In the case of Hexachlorobenzene, no pesticide was detected in the sample prepared by SPE and 0.019 ppm was detected in the sample prepared with the eXtreme Filter Vial. The use of Thomson eXtreme 0.45µm, PTFE Filter Vials as a substitute for SPE conforms to USP Method 561.

The results show Thomson eXtreme Filter Vials offer a viable alternative with higher recovery and less preparation time compared to SPE for the preparation of juices prior to pesticide analysis.

Restek or its products are not affiliated with Thomson Instrument Company



SUPPLEMENT ANALYSIS OF HUPERZINE A BY HPLC

.45µm eXtreme|FV Nylon

Huperzine A Summary

- 1. Samples are extracted with 10mM HCl (aqueous)
- 2. Non-soluble plant parts or excipients are filtered out using a $0.45 \mu m$ Nylon filter
- 3. Samples are injected onto the HPLC System

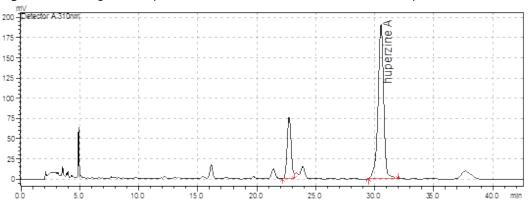
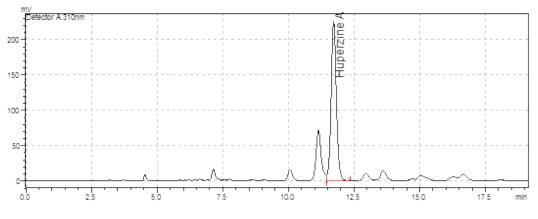


Figure I: Chromatogram of Huperzine A extracted from the Chinese Club Moss, Huperzia serrata

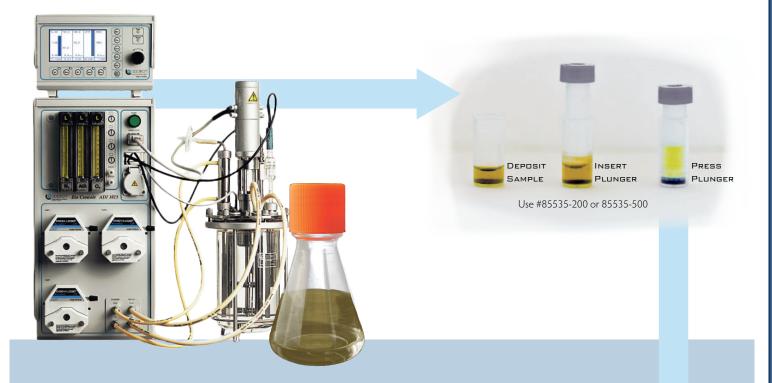
Figure II: Chromatogram of Huperzine A extracted from a Club Moss Powdered Extract







Antibody Analysis with eXtreme FV®



HPLC Column and Method

Column

Poros® Protein A by Applied Biosystem® 2-1001-00 Column

Method

A Solvent: PBS pH 7.4 B Solvent: 150 milimolar Sodium Cloride pH 2.2 Isocratic 6 minute run on an Agilent[®] 1200

Filter Vials Allow

- Real Time Monitoring
- Quantify Antibodies
- Ideal For Timepoints
- Accurate On The Fly Adjustments
- Fits In Standard Autosampler



Thomson Instrument Company is not affiliated with Agilent Technologies[®], Corning Life Sciences[®], Applied Biosystem[®] a part of Life Technologies[®] or any of their products.



ANALYSIS OF NITROSAMINES IN TOBACCO

Prep:

1. 0.25g of unburned/smokeless tobacco sample

2. Extracted with 100mM ammonium acetate solution, filtered with eXtreme|FV $^\circ$ PVDF 0.45 μ m

HPLC:

Injection Volume:	5µL
Column:	Waters Xterra MS C18, 50x4.6mm, 5µm
Aqueous phase:	5mM ammonium acetate in HPLC water
Organic Phase:	5mM ammonium acetate in 95/5 acetonitrile/water blend.

Gradient:

Time [min] Organic %

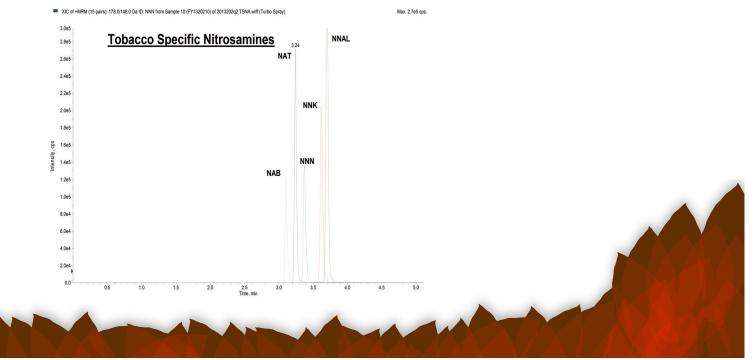
0	5
1	5
2	35
2 5	35
6	5
6 8	5

Flow rate:	1mL/min
Temperature:	60°C
Detection:	MS/MS

Analyte lo	n pair Q1/Q3 (amu)
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NAB	192/162
NAT	190/160
NNK	208/122
NNN	178/148
NNAL	210/180

N-Nitrosoanabasine N-Nitrosoanatabine N-Nitrosonornicotine 4-(Methylnitrosamino)-1-(3-pyridyl)-1-butanone 4-(Methylnitrosamino)-1-(3-pyridyl)-1-butanol





When Every µL Counts

VOLUME 10µ

MINIMUM

VOLUME 250µL

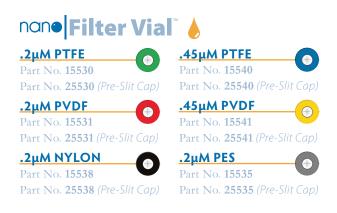
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nano|Filter Vials (10µL Dead Volume)

Patented

Thomson nano|Filter Vials[™] offer a very low dead volume allowing one to filter as little as 10µL of sample with enough remaining filtrate to make a 2µL injection. The filter vial consists of two parts: a filter vial shell with mating bottom surface and a plunger which includes a filter on one end and a screw cap vial on the other end.

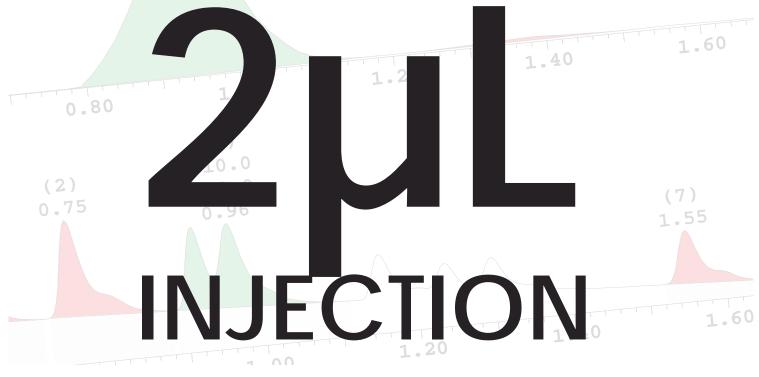
Applications include the analysis of enzymes, peptides, DNA, RNA, synthesis reaction intermediates, finished products, saliva, samples available in low volumes, in-vial evaporation and re-suspension for sample concentration and buffer/solvent change.





The strument: ZQ12 100UL (3); 310.0; FILTRATION

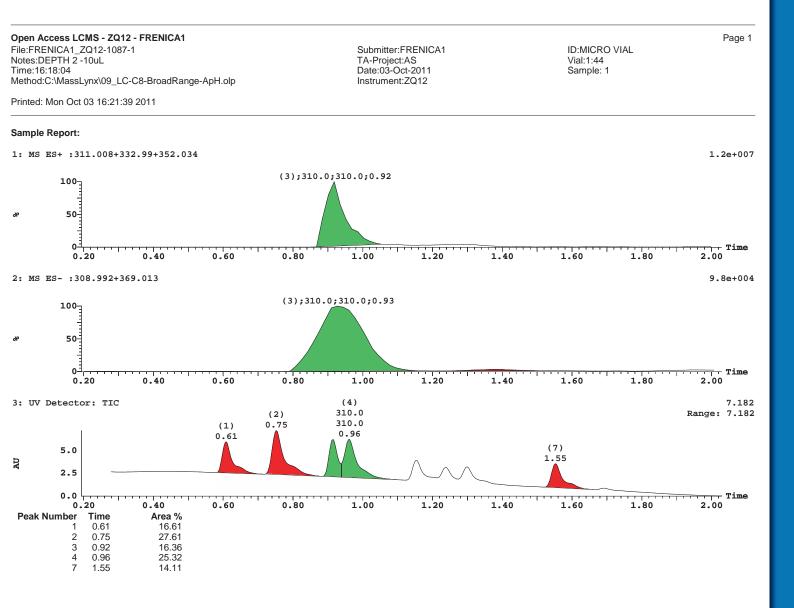
















Open Access LCMS - ZQ12 - FRENICA1 Page 2 File:FRENICA1_ZQ12-1087-1 Notes:DEPTH 2 -10uL Submitter:FRENICA1 ID:MICRO VIAL TA-Project:AS Vial:1:44 Time:16:18:04 Date:03-Oct-2011 Sample: 1 Method:C:\MassLynx\09_LC-C8-BroadRange-ApH.olp Instrument:ZQ12 Printed: Mon Oct 03 16:21:39 2011 Sample Report (continued): 3: UV Detector: 214 7.517e-1 (3) (4) 310.0310.0 Range: 7.517e-1 (1) 0.61 (2) 310.0310.0 7.5e-1-0.75 0.92 0.96 (5) 5.0e-1 1.30 PA D 2.5e-1 2.00 Time 0.0-...... 0.40 1.80 0.80 1.20 1.60 0.60 1.00 1.40 0.20 Peak Number Area % Time 28.93 0.61 1 2 0.75 19.58 3 0.92 17.71 17 56 4 5 0.96 1.30 16.23 Peak ID Time Mass Found Base Peak State 0.61 195.2 Diversity fail 1 1:(Time: 0.61) 1:MS ES+ 1.7e+006 195.2 100₇ -0 ----- m/z 600.0 1400.0 400.0 800.0 1000.0 1200.0 1600.0 1800.0 200.0 Peak ID Time Mass Found Base Peak State 1 0.61 478.1 Diversity fail 1:(Time: 0.61) 2:MS ES-1.4e+003 100 151.3 231.4 282.3 478.1 1350.4 1495.4 606.5 644.9 799.9824.4907.3 1004.3 1056.71203.6 1737.6 ^{1948.5}1996.4



Appendix

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	Housing Materials	Filter Materials			
	РР	PTFE	PVDF	PES	nylon
Acetic Acid (glacial) acid, organic	TST	R	R	R	NR
Acetone ketone	R	R	NR	GNR	R
Acetonitrile (ACN) nitrile	R	R	LTD	NR	R
Alconox, 1% surfactant/detergent	ND	TST	TST	ND	TST
Ammonium Hydroxide caustic	TST	GR	R	NR	TST
Ammonium Sulfate (saturated) salt, aqueous solution	R	GR	NR	ND	R
Amyl Acetate ester	TST	R	R	GR	TST
Amyl Alcohol alcohol	R	R	R	GR	TST
Benzene HC, aromatic	NR	R	R	NR	R
Benzyl Alcohol HC aromatic/alcohol	NR	R	R	ND	TST
Boric Acid (aqueous solution) acid, inorganic	R	GR	TST	GR	R
Butyl Acetate ester	TST	GR	TST	GNR	R
Butyl Alcohol alcohol	R	GR	R	GR	R
Carbon Tetrachloride HC, halogenated	NR	GR	R	GNR	TST
Cellosolve (Ethyl) glycol ether	R	GR	ND	GR	R
CHAPS (aqueous solution) surfactant/detergent	ND	TST	ND	ND	TST
Chloroform HC, halogenated	NR	R	R	GNR	NR
Cyclohexanone ketone	NR	R	NR	GNR	R
Diethyl Pyrocarbonate, 0.2% carboxylic anhydride	ND	ND	TST	ND	ND
Dimethyl Sulfoxide (DMSO) sulfoxide	R	R	NR	NR	R
Dimethylacetamide amide	R	GR	NR	NR	NR
Dimethylformamide amide	R	GR	NR	ND	R
Dioxane ether	R	GR	R	ND	R

Key to the Above Table

ND = No Data Presently Available

= Recommended = Generally Recommended

R

GR

NR = Not Recommended GNR = Generally Not Recommended **TST** = Testing Recommended LTD = Limited Recommendation

	Housing Materials	Filter Materials			
	РР	PTFE	PVDF	PES	nylon
Ethers ether	NR	R	R	ND	R
Ethyl Acetate ester	TST	R	R	GNR	R
Ethyl Alcohol alcohol	R	R	R	GR	TST
Ethylene Glycol glycol	R	R	R	GR	R
Formaldehyde aldehyde	R	R	R	ND	R
Formic Acid, 50% acid, organic	R	GR	R	ND	NR
Freon (TF or PCA) HC, halogenated	R	GR	R	ND	R
Gasoline HC	NR	R	R	GR	R
Glycerine (Glycerol) glycol	R	GR	R	GR	R
Guanidine Hydrochloride, 6M salt, aqueous solution	ND	GR	ND	ND	ND
Guanidine Thiocyanate, 5M salt, aqueous solution	ND	GR	ND	ND	ND
Helium gas	R	R	TST	ND	R
Hexane HC, aliphatic	NR	R	R	GR	R
Hydrochloric Acid, 1N (HCL) acid, inorganic	GR	R	R	GR	GR
Hydrochloric Acid, 6N (HCL) acid, inorganic	TST	R	TST	GR	TST
Hydrochloric Acid, conc. (HCL) acid, inorganic	NR	R	NR	ND	NR
Hydrofluoric Acid acid, inorganic	NR	R	NR	NR	NR
Hydrogen gas	R	R	R	ND	R
Hydrogen Peroxide, 3% peroxide	R	R	R	ND	R
Hydrogen Peroxide, 30% peroxide	TST	R	R	ND	TST
Hydrogen Peroxide, 90% peroxide	R	R	R	ND	NR
HYPO (aqueous solution) salt, aqueous solution	R	GR	R	ND	R
Isobutyl Alcohol alcohol	R	R	R	GR	TST

Key to the Above Table **ND** = No Data Presently Available

= Recommended GR = Generally Recommended

R

NR = Not Recommended **GNR** = Generally Not Recommended **TST** = Testing Recommended **LTD** = Limited Recommendation

	Housing Materials	Filter Materials			
	РР	PTFE	PVDF	PES	nylon
Isopropyl Acetate ester	TST	R	R	GNR	R
Isopropyl Alcohol alcohol	R	R	R	GR	TST
Kerosene HC	TST	LTD	R	GR	R
Lactic Acid, 50% acid, organic/alcohol	R	GR	TST	ND	TST
Lubrol PX (aqueous solution) surfactant/detergent	ND	TST	ND	ND	ND
Methyl Ethyl Ketone (MEK) ketone	R	R	NR	GNR	R
Mercaptoethanol, 0.1M alcohol/mercaptan	ND	ND	ND	ND	ND
Methyl Acetate ester	TST	R	NR	GNR	R
Methyl Alcohol alcohol	R	R	R	GR	TST
Methylene Chloride HC, halogenated	NR	R	NR	GNR	TST
Methyl Isobutyl Ketone (MIBK) ketone	NR	R	NR	GNR	R
Mineral Spirits HC	NR	R	R	GR	R
Nitric Acid, 6N acid, inorganic	TST	R	R	R	NR
Nitric Acid (concentrated) acid, inorganic	NR	ND	NR	ND	NR
Nitrobenzene HC, aromatic	NR	R	R	ND	R
Nitrogen gas	ND	R	R	ND	R
Nonidet-P40 (aqueous solution) surfactant/detergent	ND	ND	ND	ND	ND
Ozone gas	NR	GR	R	ND	NR
Paraldehyde aldehyde	TST	GR	TST	ND	R
Pentane HC, aliphatic	NR	GR	GR	GR	R
Petroleum Ether ether	ND	GR	R	ND	R
Phenol (aqueous solution) phenol	NR	GR	R	ND	NR
Potassium Hydroxide, 3N caustic	R	R	R	ND	R

Key to the Above Table

ND = No Data Presently Available

Recommended
 Generally Recommended

R

GR

NR = Not Recommended

TST = Testing Recommended

38

GNR = Generally Not Recommended

LTD = Limited Recommendation

	Housing Materials	Filter Materials			
	PP	PTFE	PVDF	PES	nylon
Pyridine amine	R	GR	NR	NR	TST
Silicone Oils silicone	R	GR	R	ND	R
Sodium Carbonate (aqueous solution) salt, aqueous solution	R	R	R	ND	TST
Water (Brine) salt, aqueous solution	R	R	R	ND	R
Sodium Chloride (aqueous solution) salt, aqueous solution	R	R	R	ND	R
Sodium Dodecyl Sulfate surfactant/detergent	ND	ND	ND	ND	ND
Sodium Hydroxide, 3N caustic	R	R	R	R	R
Sodium Hydroxide (concentrated) caustic	R	R	R	R	NR
Sulfuric Acid (concentrated) acid, inorganic	NR	R	TST	GNR	NR
Tetrahydrofuran (THF) ether	NR	GR	NR	ND	R
Toluene HC, aromatic	NR	R	R	R	R
TCA (aqueous solution) acid, organic	R	GR	R	ND	TST
Trichloroethane HC, halogenated	NR	R	TST	GNR	TST
Trichloroethylene HC, halogenated	NR	R	R	GNR	TST
Tween 20 (aqueous solution) surfactant/detergent	ND	R	TST	ND	TST
Urea, 8M salt, aqueous solution	R	GR	R	ND	R
Xylene HC, aromatic	NR	R	R	ND	R

ND = No Data Presently Available GR = Generally Recommended GNR = Generally Not Recommended LTD = Limited Recommendation

Key to the Above Table R = Recommended NR = Not Recommended TST = Testing Recommended

Compound Compatibility

	PVDF	PES	PTFE	PES	PVDF
Filter to be used	.2µm	.2µm	.2µm	.45µm	.45µm
5-Fluorouracil					
(18F) Fluoromisondazole, Misiomidazole					
Acebutolol		•			
Acetylsalicylic acid		0			
Alpha1-Proteinase Inhibitor (Human)					0
Alprenolol		0			
Amiloride		•			
Amphotericin B for Injection USP					0
Atenolol		•			
Azathioprine				0	0
Azodicarbonamide		•			
Bleomycin Sulfate			0		
Caffeine		•			
Cetirizine				0	0
Chlorothiazide		•			
Chloramphenicol		0			
Cimetidine					
Ciprofloxacin		0			
Cisplatin, Cisplatin Injection					
Cyclosporine A					
Cytarabine					
Daunorubicin			•		
DE-310					
Diclofenac					
Enalapril					
Ethionamide					
Factor IX Complex Heat-Treated					
Gatifloxacin				0	0
Hydrochlorothiazide					

For more information and the references to this table please see htslabs.com.

Compound Compatibility

Filter to be used	PVDF	PES	PTFE	PES	PVDF
Filter to be used	.2µm	.2µm	.2µm	.45µm	.45µm
Ibuprofen				0	
losniazid			•		
isonicotinic acid			0		
Ketamine		•			
Las 35917					0
Levofloxacin					
Lomefloxacin				0	0
Methyl Gag; NSC-32946					
Metoprolol		0			
Mitomycin					
Morphazinamide					
Nadolol					
Nicotinic acid					
Paclitaxel					
p-Aminobenzoic acid (PABA)					0
p-aminosalicylic acid					
Pefloxacin					0
Pentoxifylline (PTX)					
Phenytoin					0
Pyrazinamide					
Pyrimethamine					0
Ranitidine					
Rifampicin					0
Sabeluzole					
Streptokinase					0
Sulfadozine					
Sulphasalazine					
Sulpiride					
Terbutaline					

For more information and the references to this table please see htslabs.com.

Compound Compatibility

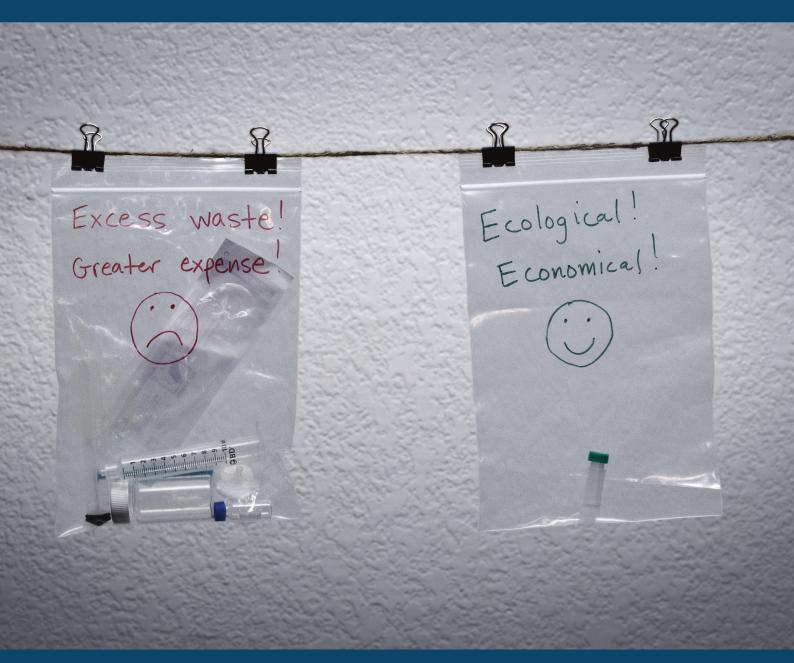
Filter to be used	PVDF	PES	PTFE	PES	PVDF
	.2µm	.2µm	.2µm	.45µm	.45µm
Thiotepa Parenteral Sterile					
Timolol		0			
Tobramycin Vincristine Sulfate					
Tranexamic acid		0			
Triamcinolone Acetonide		•			
Triazinate; NSC-139105			•		
Tropicamide				•	
Vinblastine Sulfate					

For more information and the references to this table please see htslabs.com.

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